

A Review of Platelet-Rich Plasma: History, Biology, Mechanism of Action, and Classification

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Keywords

Platelet-rich plasma · Plasma · Autologous therapy · Biology · Mechanism of action

Abstract

Platelet-rich plasma (PRP) is currently used in different medical fields. The interest in the application of PRP in dermatology has recently increased. It is being used in several different applications as in tissue regeneration, wound healing, scar revision, skin rejuvenating effects, and alopecia. PRP is a biological product defined as a portion of the plasma fraction of autologous blood with a platelet concentration above the baseline. It is obtained from the blood of patients collected before centrifugation. The knowledge of the biology, mechanism of action, and classification of the PRP should help clinicians better understand this new therapy and to easily sort and interpret the data available in the literature regarding PRP. In this review, we try to provide useful information for a better understanding of what should and should not be treated with PRP.

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History of Platelet-Rich Plasma

Platelet-rich plasma (PRP) is also known as platelet-rich growth factors (GFs), platelet-rich fibrin (PRF) matrix, PRF, and platelet concentrate.

The concept and description of PRP started in the field of hematology [1]. Hematologists created the term PRP in the 1970s in order to describe the plasma with a platelet count above that of peripheral blood, which was initially used as a transfusion product to treat patients with thrombocytopenia [2].

Ten years later, PRP started to be used in maxillofacial surgery as PRF. Fibrin had the potential for adherence and homeostatic properties, and PRP with its anti-inflammatory characteristics stimulated cell proliferation [3].

Subsequently, PRP has been used predominantly in the musculoskeletal field in sports injuries. With its use in professional sportspersons, it has attracted widespread attention in the media and has been extensively used in this field [4]. Other medical fields that also use PRP are cardiac surgery, pediatric surgery, gynecology, urology, plastic surgery, and ophthalmology [5].

More recently, the interest in the application of PRP in dermatology; i.e., in tissue regeneration, wound healing, scar revision, skin rejuvenating effects, and alopecia, has increased [1, 6–14].

Wounds have a proinflammatory biochemical environment that impairs healing in chronic ulcers. In addition, it is characterized by a high protease activity, which decreases the effective GF concentration. PRP is used as an interesting alternative treatment for recalcitrant wounds because it is a source of GFs and consequently has mitogenic, angiogenic, and chemotactic properties [12].

In cosmetic dermatology, a study performed *in vitro* demonstrated that PRP can stimulate human dermal fibroblast proliferation and increase type I collagen synthesis [13]. Additionally, based on histological evidence, PRP injected in human deep dermis and immediate subdermis induces soft-tissue augmentation, activation of fibroblasts, and new collagen deposition, as well as new blood vessels and adipose tissue formation [14, 15].

Another application of PRP is the improvement of burn scars, postsurgical scars, and acne scars [16]. According to the few articles available, PRP alone or in combination with other techniques seems to improve the quality of the skin and leads to an increase in collagen and elastic fibers.

In 2006, PRP has started to be considered a potential therapeutic tool for promoting hair growth and has been postulated as a new therapy for alopecia, in both androgenetic alopecia and alopecia areata. Several studies have been published that refer to the positive effect PRP has on androgenetic alopecia, although a recent meta-analysis suggested the lack of randomized controlled trials [17]. As stated by the authors, controlled clinical trials are considered the best way to provide scientific evidence for a treatment and avoid potential bias when assessing efficacy [18].

Platelet Biology

All blood cells derive from a common pluripotent stem cell, which differentiates into different cell lines. Each of these cell series contains precursors that can divide and mature.

Platelets, also called thrombocytes, develop from the bone marrow. Platelets are nucleated, discoid cellular elements with different sizes and a density of approximately 2 μm in diameter, the smallest density of all blood cells. The physiological count of platelets circulating in the

blood stream ranges from 150,000 to 400,000 platelets per μL .

Platelets contain several secretory granules that are crucial to platelet function. There are 3 types of granules: dense granules, α -granules, and lysosomes. In each platelet there are approximately 50–80 granules, the most abundant of the 3 types of granules.

Platelets are primarily responsible for the aggregation process. The main function is to contribute to homeostasis through 3 processes: adhesion, activation, and aggregation. During a vascular lesion, platelets are activated, and their granules release factors that promote coagulation [19].

Platelets were thought to have only hemostatic activity, although in recent years, scientific research and technology has provided a new perspective on platelets and their functions. Studies suggest that platelets contain an abundance of GFs and cytokines that can affect inflammation, angiogenesis, stem cell migration, and cell proliferation [20].

PRP is a natural source of signaling molecules, and upon activation of platelets in PRP, the P-granules are degranulated and release the GFs and cytokines that will modify the pericellular microenvironment. Some of the most important GFs released by platelets in PRP include vascular endothelial GF, fibroblast GF (FGF), platelet-derived GF, epidermal GF, hepatocyte GF, insulin-like GF 1, 2 (IGF-1, IGF-2), matrix metalloproteinases 2, 9, and interleukin 8 [1, 21].

Definition

PRP is a biological product defined as a portion of the plasma fraction of autologous blood with a platelet concentration above the baseline (before centrifugation) [22]. As such, PRP contains not only a high level of platelets but also the full complement of clotting factors, the latter typically remaining at their normal, physiologic levels [23]. It is enriched by a range of GFs, chemokines, cytokines, and other plasma proteins [4].

The PRP is obtained from the blood of patients before centrifugation. After centrifugation and according to their different density gradients, the separation of blood components (red blood cells, PRP, and platelet-poor plasma [PPP]) follows.

In PRP, besides the higher concentration of platelets, other parameters need to be taken into account, such as the presence or absence of leucocytes and activation. This will define the type of PRP used in different pathologies.

There are several commercial devices available, which simplify the preparation of PRP. According to the manufacturers, PRP devices usually achieve a concentration of PRP 2–5 times the baseline concentration. Although one might think that a higher platelet count with a higher number of GFs would lead to better results, this has not been determined yet [20]. In addition, 1 study also suggests that a concentration of PRP 2.5 times above the baseline could have an inhibitory effect [24].

Mechanism of Action in Alopecia

The GFs and the bioactive molecules present in PRP promote 4 main actions in the local environment of the administration, such as proliferation, migration, cell differentiation, and angiogenesis [22, 25–28]. Various cytokines and GFs are involved in the regulation of hair morphogenesis and cycle hair growth [29].

The dermal papilla (DP) cells produce GFs such as IGF-1, FGF-7, hepatocyte growth factor, and vascular endothelial growth factor that are responsible for maintaining the hair follicle in the anagen phase of the hair cycle. Therefore, a potential target would be to upregulate these GFs within the DP cells, which lengthen the anagen phase [30].

According to a study performed by Akiyama et al. [31], epidermal growth factor and transforming growth factor are involved in regulating the growth and differentiation of bulge cells, and platelet-derived growth factor may have related functions in the interactions between the bulge and the associated tissues, starting with follicle morphogenesis [27].

Beside the GFs, the anagen phase is also activated by Wnt/ β -catenin/T-cell factor lymphoid enhancer [32]. In the DP cells, the activation of Wnt will lead to an accumulation of β -catenin, which, in combination with the T-cell factor lymphoid enhancer, also acts as a co-activator of transcription and promotes proliferation, survival, and angiogenesis. The DP cells then initiate the differentiation and consequently the transition from the telogen to the anagen phase [25]. β -Catenin signaling is important in human follicle development and for the hair growth cycle.

Another pathway presented in DP is the activation of extracellular signal-regulated kinase (ERK) and protein kinase B (Akt) signaling that promotes cell survival and prevents apoptosis [6].

The precise mechanism by which PRP promotes hair growth is not fully understood. To explore the possible

mechanisms involved, Li et al. [6] performed a well-designed study to investigate the effects of PRP on hair growth using in vitro and in vivo models. In the in vitro model, activated PRP was applied to human DP cells obtained from normal human scalp skin. The results demonstrated that PRP increased the proliferation of human DP cells by activating ERK and Akt signaling, leading to antiapoptotic effects. PRP also increased the β -catenin activity and FGF-7 expression in DP cells. Concerning the in vivo model, mice injected with activated PRP showed a faster telogen-to-anagen transition in comparison to the control group.

Recently, Gupta and Carviel [25] also proposed a mechanism for the action of PRP on the human follicles that includes the “elicitation of the Wnt/ β -catenin, ERK, and Akt signaling pathways fostering cell survival, proliferation, and differentiation.”

After GF binds with its correspondent GF receptor, the signaling necessary for its expression begins. The GF-GF receptor activates the expression of both Akt and ERK signaling. The activation of Akt will inhibit 2 pathways through phosphorylation: (1) the glycogen synthase kinase-3 β that promotes degradation of β -catenin, and (2) Bcl-2-associated death promoter, which is responsible for inducing apoptosis. As stated by the authors, PRP might increase vascularization, prevent apoptosis, and prolong the duration of the anagen phase [27].

The main functions of the GFs presented in the PRP are detailed in Table 1.

Devices to Obtain PRP

Currently, there is a great discussion and no consensus regarding PRP preparation. PRP is prepared through a process known as differential centrifugation, in which acceleration force is adjusted to sediment certain cellular constituents based on different specific gravity [33].

Regarding the preparation of PRP, there are 2 techniques:

1. Open technique: the product is exposed to the environment of the working area and comes in contact with different materials that should be used for their production, such as pipettes or product-collection tubes. In the blood processing to obtain PRP with the open technique, it should be guaranteed that the product is not contaminated during microbiological handling.
2. Closed technique: it involves the use of commercial devices with CE marking (including centrifuge equip-

ment and application) in which the product is not exposed to the environment (recommended).

Several CE medical devices are available for the production of autologous PRP. Most of them are included in one of the following 3 types of devices:

1. The blood is obtained with a tube that contains an anticoagulant, and this tube can be used for any type of centrifuge.
2. Medical devices with which the blood is collected into a tube that already contains an anticoagulant; the centrifugation can then be made in any type of centrifuge.
3. Medical devices with which the blood is collected into a syringe previously filled with an anticoagulant; usually, the blood is transferred into a secondary device whose shape imposes the use of the centrifuge supplied by the same manufacturer [34].

The preparation of PRP depends on the type of device chosen and should be done according to the manufacturer's instructions (Table 2).

As described in Table 2, there are different PRP systems that facilitate the preparation of PRP in a reproducible manner. All operate on a small volume of drawn blood and on the principle of centrifugation.

Briefly, the procedure requires the use of relatively small volumes of blood. The PRP is obtained from the blood of patients before centrifugation. The whole blood is obtained by venipuncture in anticoagulated tubes (usually with acid citrate dextrose or sodium citrate solution). The blood is then centrifuged with single- or a double-spin centrifugation, depending on the device. The settings of the centrifuge established to obtain PRP at an adjustable concentration are defined by the manufacturer and cannot be changed by the physician.

After centrifugation, the tube shows 3 basic layers: at the bottom of the tube, there are red blood cells with leukocytes deposited immediately above; the middle layer corresponds to the PRP, and at the top, there is the PPP (Fig. 1). The PPP is removed, and PRP is obtained. Platelets can be activated before application of the PRP, although there is no consensus on whether or not platelets must be previously activated before their application and with which agonist [35].

Thrombin and calcium chloride, which are aggregation inducers, are used with the aim to activate platelets and stimulate degranulation, causing the release of the GFs [36].

Some authors activate platelets, whereas others apply platelets without previously activating them, arguing that better results are obtained. Recent studies found that the use of such aggregators is not necessary because at the

Table 1. Main functions of the growth factors present in platelet-rich plasma

Growth factors	Main functions
PDGF [27]	<ul style="list-style-type: none"> - Increases hair growth - Vascularization - Angiogenesis stimulator
TGF- β [29]	<ul style="list-style-type: none"> - Inhibits hair growth in vitro - Hair-cell proliferation and regeneration
VEGF [28]	<ul style="list-style-type: none"> - Expressed in DP cells in the anagen phase - Probably regulates perifollicular angiogenesis - Increases perifollicular vessel size during the anagen growth phase
EGF [29, 43]	<ul style="list-style-type: none"> - Angiogenesis stimulator - Hair-cell proliferation and regeneration
HGF [36]	<ul style="list-style-type: none"> - Angiogenesis stimulator
FGF [29, 43–45]	<ul style="list-style-type: none"> - Increases hair growth by inducing the anagen phase of HF - Promotes DP cell proliferation - Increases the HF size in mice - Angiogenesis stimulators
IGF-1 [6, 27, 46]	<ul style="list-style-type: none"> - Increases hair growth - Maintains HF growth in vitro - Angiogenesis stimulator

PDGF, platelet-derived growth factor; TGF, transforming growth factor; VEGF, vascular endothelial growth factor; DP, dermal papilla; EGF, epidermal growth factor; HGF, hepatocyte growth factor; FGF, fibroblast growth factor; IGF-1, insulin-like growth factor 1; HF, human follicle(s).

time of administration, the platelets are automatically released and ready to exert their function.

Even though most devices aim to obtain the best PRP, the systems differ extensively in their ability to collect and concentrate platelets depending on the method and time of its centrifugation. As a result, suspensions of different concentrations of platelets and leukocytes are obtained. It is difficult to assess which kit for PRP preparation is better and which is worse [33]. In addition, each preparation may produce different types of PRP with different applications. There is no consensus about the number of centrifugations required, nor on their speed duration.

There is intensive ongoing debate regarding the ideal volume of PRP to administer, the frequency of application, the exact site of administration of PRP, and which technique/preparation system of manufacture to use.

Table 2. Blood collection and centrifugation protocols from different medical devices to obtain platelet-rich plasma

Devices	Blood collection/ anticoagulant	Centrifugation		
		number of times	speed/time	centrifuge
Selphyl®	Tube 9 mL/sodium citrate	1	1,100 g/6 min	Classic
PRGF Endoret®	Tube 9 mL/sodium citrate	1	270 g/7 min	Classic
Cascade®	Tube 9 mL/sodium citrate	2	1,100 g/6 min 1,450 g/15 min	Classic
Plateltex®	Tube 9 mL/ACD	2	180 g/10 min 1,000 g/10 min	Classic
Regenkit®	Tube 9 mL/sodium citrate	1	1,500 g/9 min	Classic
ACP Arthrex®	Syringe 15 mL/ACD or no anticoagulant	1	1,500 rpm/5 min	Adapted
GPS III®	Syringe 30 or 60 mL/ACD	1	3,200 rpm/15 min	Adapted
Genesis®	Syringe 12 mL/ACD	1	2,400 rpm/12 min	Adapted
SmartPrep 2®	Syringe 20 or 60 mL/ACD	2	2,500 rpm/4 min 2,300 rpm/10 min	Adapted
Proteal®	Syringe 20 mL/sodium citrate	1	1,800 rpm/8 min	Adapted
Magellan®	Syringe 30–60 mL/ACD	–	–	Adapted device
Angel®	Syringe 40–180 mL/ACD	–	–	Adapted device

ACD, acid citrate dextrose. Courtesy of Dr. Jeremy Magalon, adapted from Dohan Ehrenfest [38].

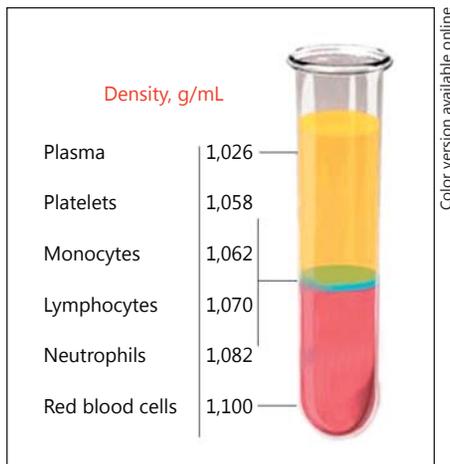


Fig. 1. After centrifugation, the blood components (red blood cells, leukocytes, and platelets) are separated from the plasma due to their different densities. The platelets have the lowest density. Adapted from Dohan Ehrenfest et al. [38].

Classification

Several authors have tried to characterize and classify the numerous techniques available on the market in terms of preparation (centrifugation speed and use of antico-

agulant), content (platelets, leucocytes, and GFs), and applications [36].

Although the literature about PRPs developed with all these contradictions, the need for a standardized terminology is of great importance [36, 37]. Thus, some classifications have been proposed to achieve a consensus terminology in the field of platelet concentrates [37–42].

Characterizing the type of PRP used (as a pure PRP in our study) will lead to a better understanding of PRP, and data available will be easier to sort and interpret. Furthermore, this terminology would serve as a basis for further research on the topic.

In 2009, Dohan Ehrenfest et al. [38] proposed a classification of 4 main families of preparations following 2 principle parameters: presence or absence of a cell content (such as leucocytes) and the fibrin architecture:

1. Pure PRP or leucocyte-poor PRP: the preparation obtained is without leucocytes and shows a low-density fibrin network after activation.
2. Leucocyte and PRP: the preparations contain leucocytes and show a low-density fibrin network after activation.
3. Pure PRF or leucocyte-poor PRF: preparations are without leucocytes and with a high-density fibrin network. Unlike pure PRP or PRP containing leucocytes,

these products cannot be injected and exist in an activated gel form.

4. Leucocyte-rich fibrin and PRF: products are preparations with leucocytes and with a high-density fibrin network.

Mishra et al. [40] proposed another classification based on the presence or absence of white blood cells, activation status, and platelet concentration, based on the coefficients of an increase in the platelet and leukocyte concentration in PRP compared to the whole-blood baseline, as well as on PRP activation [41].

The classifications were not consensual and there is still the intent to search a classification for PRP that could characterize the injected PRP in order to compare the efficacy of different studies.

An important point of discussion is that in the previous classifications, the authors did not take into account the final volume of the preparation, the presence or absence of red blood cells (RBCs) in PRP, and the doses of platelets in the final volume of PRP obtained.

In 2016, Magalon et al. [41] proposed the DEPA classification (Dose, Efficiency, Purity, Activation) that focuses on the quantity of platelets obtained by the PRP kits as well as on product purity and on platelet activation prior to injection.

The DEPA classification is based on 4 different components:

1. Dose of injected platelets: calculated by multiplying the platelet concentration in PRP by the obtained volume of PRP. According to the injected dose (measured in billions or millions of platelets), it should be categorized into (a) very high dose of injected platelets of >5 billion; (b) high dose of injected platelets, from 3 to 5 billion; (c) medium dose of injected platelets, from 1 to 3 billion, and (d) low dose of injected platelets, <1 billion.
2. Efficiency of the production: corresponds to the percentage of platelets recovered in the PRP from the blood. It is categorized as follows: (a) high device efficiency, if the recovery rate in platelets is >90%; (b) medium device efficiency, if the recovery rate in platelets is between 70 and 90%; (c) low device efficiency, if the recovery rate is between 30 and 70%, and (d) poor device efficiency, if the recovery rate is <30% and corresponds to the relative composition of platelets, leucocytes, and RBCs in the obtained PRP.
3. Purity of the PRP obtained: correlates to the relative composition of platelets, leucocytes, and RBCs in the obtained PRP. It is described as (a) very pure PRP, if the percentage of platelets in the PRP, compared with RBCs and leucocytes, is >90%; (b) pure PRP, between

70 and 90% of the platelets; (c) heterogeneous PRP, if the percentage of platelets is between 30 and 70%, and (d) whole-blood PRP, if the percentage of platelets in the PRP is <30% compared with RBCs and leucocytes.

4. Activation process: if an exogenous clotting factor was used to activate platelets, such as autologous thrombin or calcium chloride.

Although this last classification is very complete, this quantification cannot be defined by the physician and should be registered in each CE medical device available for preparation of PRP.

Conclusion

PRP is being used as a new therapeutic option for different pathologies in the field of dermatology, such as trichology, wound healing, and cosmetic medicine. In this manner, understanding the biology and mechanism of action of this therapy should help clinicians in selecting a system that meets their specific needs for a given indication. In addition, characterizing the type of PRP used will lead to a standardization of PRP, making it easier to sort and interpret available data. We hope that this review serves as a basis for further research on the use of PRP.

Disclosure Statement

All authors have no conflicts of interest to disclose.

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